A cross-sectional study of the histopathology and immunology of alopecia areata: Unearthing the role of the Janus kinase—signal transducer and activator of transcription pathway

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Abstract

Background: Alopecia areata is an autoimmune disease that occurs as a result of the loss of the inherent immune privilege of the hair follicle. It has been recently demonstrated that the interferon-γ/interleukin-15 feedback loop that signals via the Janus kinase–signal transducer and activator of transcription pathway is critical to the breakdown of this immune privilege.

Aims: To evaluate the immunological distribution of CD4+ T-cells, CD8+ T-cells, phosphorylated signal transducer and activator of transcription 1 and study its relation with the clinical and histopathological findings of the disease.

Materials and Methods: A total of 30 patients of alopecia areata were included in the study. Following a detailed history and clinical examination, a scalp biopsy was performed. Histopathology was studied and immunohistochemistry was done to demonstrate the positivity and distribution of CD4+ T-cells, CD8+ T-cells and phosphorylated signal transducer and activator of transcription 1.

Results: The follicular count, number of anagen and terminal hair were found to be decreased, whereas the catagen, telogen and vellus hair were found to be increased in number. A peribulbar CD4+ T-cell infiltrate was seen in 70% cases, whereas a CD8+ T-cell infiltrate was seen in 83.3% cases. An intrabulbar CD4+ T-cell infiltrate was seen in 26.7% cases, whereas a CD8+ T-cell infiltrate was seen in 70% cases. Among the 25 hair follicles dermal papilla identified, 36.8% cases were found to be positive for phospho-signal transducer and activation of transcription-1.

Limitations: The drawbacks of our study included a small sample size and the use of only vertical sectioning for the scalp biopsy samples.

Conclusion: Phosphorylated signal transducer and activator of transcription 1 positivity as an indicator of signalling via the Janus kinase-1/2 pathway was seen in 36.8% of our cases highlighting the integral role of this pathway in the pathogenesis of alopecia areata.

Key words: Alopecia areata, histopathology, immunology, Janus kinase–signal transducer and activator of transcription, pathogenesis

Access this article online							
Quick Response Code:	Website: www.ijdvl.com						
	DOI: 10.4103/ijdvl.IJDVL_738_17						

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How to cite this article: Bansal A, Relhan V, Garg VK, Saran RK. A cross-sectional study of the histopathology and immunology of alopecia areata: Unearthing the role of the Janus kinase–signal transducer and activator of transcription pathway. Indian J Dermatol Venereol Leprol 2019;85:455-61.

Received: December, 2017. Accepted: September, 2018.

Introduction

Alopecia areata is a noncicatricial alopecia with an incidence of 0.1-0.2% in the world with an equal preponderance in males and females.^{1,2} It is most commonly reported on the scalp, typically as a smooth, round, bald patch, while the complete loss of scalp hair (alopecia totalis) or the complete loss of scalp and body hair (alopecia universalis) have also been reported.^{2,3} Alopecia areata is considered to be a hair-specific autoimmune disease, characterized by a T-cell-mediated inflammation of the anagen hair. Among genetically predisposed individuals, this happens as a result of the collapse of its immune privilege, mediated by interferon-y. This leads to major histocompatibility complex class I expression in the proximal hair bulb, thus expressing self-epitopes to cytotoxic CD8+ T-cells, which subsequently enter the hair follicle. CD4+ T-cells constitute the majority of lymphocytes in the perifollicular area. 4,5 This peribulbar and intrabulbar lymphocytic infiltrate around anagen follicles, resembling a "swarm of bees", characteristic of alopecia areata.6

Recently, the role of CD8+ natural killer, group 2, member D protein (NKG2D) T-cells and the interferon-γ/interleukin-15 feedback loop has come into the picture. These function via the Janus kinase–signal transducer and activator of transcription pathway. Interferon-γ released from these activated T-cells leads to signal transducer and activator of transcription-1 activation, causing it's translocation to the nucleus, which leads to interleukin-15 production. Interleukin-15 enhances the production of interferon-γ, thus perpetuating this critical feedback loop.

We undertook this study to elucidate the clinical and histopathological findings of this disease process and understand the role of the Janus kinase–signal transducer and activator of transcription pathway in the pathogenesis of this disease process via demonstration of phosphorylated transducer and activator of transcription-1 in the hair follicles of patients with alopecia areata. There are very few studies that have evaluated this aspect, hence we decided to undertake this study.

Materials and Methods

This was a cross-sectional study carried out between October 2015 and March 2017. Thirty patients of alopecia areata presenting to the outpatient clinic of the Department of Dermatology, Maulana Azad Medical College and associated

Lok Nayak Hospital, New Delhi were recruited. Ethical approval was obtained from Institutional Review Board vide letter IRBNo. IEC/MAMC/2015/717.

Pregnant or lactating females; patients with history of intake and/or application of corticosteroids in any form within last 1 month were excluded from the study.

A detailed history was obtained and clinical examination done and recorded on predesigned proformas. Severity of hair loss was assessed using the Severity of Alopecia Tool Score. The percentage of scalp hair loss in both sides, back and top of the scalp were determined independently. Each value was multiplied by the % scalp covered in that area of the scalp and the products of each section summed for a final total % hair loss, designated as the SALT score with the following categories: $S_0 =$ no hair loss; $S_1 \le 25\%$ hair loss; $S_2 = 25-49\%$ hair loss; $S_3 = 50-74\%$ hair loss; $S_4 = 75-99\%$ hair loss; $S_5 = 100\%$ hair loss.⁸

A biopsy was performed under local anesthesia from the margin of the lesion, using a 4 mm punch for the purpose of histopathological and immunological evaluation. The tissue was placed in 10% buffered formalin and processed for histopathology by routine paraffin embedding in a vertical orientation. Then, 3-5 µm sections were cut and stained with hematoxylin and eosin. Immunohistochemistry was done on the unstained sections using the antibodies against CD4 (monoclonal mouse antihuman CD4, DAKO, ready to use) CD8 (monoclonal mouse antihuman CD8, Thermoscientific, ready to use) and phospho-signal transducer and activation of transcription-1 (Tyr701) (58D6). Intrabulbar and peribulbar CD4+ T-cells and CD8+ T-cells were identified and the grading was done, as shown in Table 1.9,10 Phosphorylated transducer and activator of transcription-1 nuclear staining was assessed in the hair follicle dermal papilla and graded as shown in Table 1.

The primary outcome measures were the number of hair follicle dermal papillae demonstrating phosphorylated transducer and activator of transcription-1 positivity and the number of hair follicles showing CD4+/CD8+ T-cells in the peribulbar and intrabulbar location based on immunohistochemistry.

The secondary outcome measures were the relations between the immunological pattern of CD4+ T-cells, CD8+ T-cells, phosphorylated transducer and activator of transcription-1

Table 1: Immunohistochemical grading used for the lymphocytic infiltrate, CD4+, CD8+ T-cells and phosphorylated transducer and activator of transcription-1

- II					
Grading	0	1	2	3	
Lymphocytic infiltrate grade	Absent	Mild (+)	Moderate (++)	Dense (+++)	
CD4+ and CD8+ T-cell density grade	No cell	Rare cells, no cluster	Small clusters	Diffuse clusters	
Phosphorylated transducer and activator	No nuclear staining	Faint nuclear staining	Moderate nuclear staining	Intense nuclear staining	
of transcription-1 positivity grade					

with the clinical (disease duration, pattern and severity) and histopathological (lymphocytic infiltrate) parameters.

The quantitative variables were expressed as mean \pm SD and compared between groups using Kruskal–Wallis test, unpaired *t*-test and Mann–Whitney *U*-test wherever applicable. Also, qualitative variables were expressed as frequencies/percentages and compared between groups using Chi-square test and Fisher's exact test wherever applicable. Data were analyzed using Statistical Packages for the Social Sciences, and a *P* value <0.05 was considered significant.

Results

The study population comprised 30 patients, 19 males and 11 females. The distribution of age in our study ranged from 4 years to 57 years with the mean age of 24.37 ± 14.048 years. The duration of the disease in our study ranged from 2 days to 8 years, with the maximum number of patients (30%) presenting within the first 3 months of disease. In our study, 22 patients (73.3%) presented with alopecia areata, whereas 6 (20%) had alopecia universalis and 2 (6.7%) had alopecia totalis. Among the 22 patients having patchy hair loss, 4 (13.3%) had patterned hair loss. Out of these, 3 (10%) patients presented with an ophiasis pattern and 1 (3.3%) patient with a reticular pattern.

In our study, the follicular count ranged from 3 to 32, with a mean follicular count of 11.93 ± 7.235 . The mean number of anagen hair were 1.63 ± 1.586 , and the mean number of telogen hair were 7.84 ± 5.471 . Mean number of terminal hair were 7.83 ± 5.471 and mean vellus hair were 2.47 ± 2.417 . The terminal hair to vellus hair ratio was 3.8:1 and anagen to telogen hair ratio was 1:5.

A peribulbar lymphocytic infiltrate was identified in 27 (90%) of the cases [Figures 1 and 2]. A peribulbar CD4+ T-cell infiltrate was seen in 21 (70%) cases, whereas a CD8+ T-cell infiltrate was seen in 25 (83.3%) cases. An

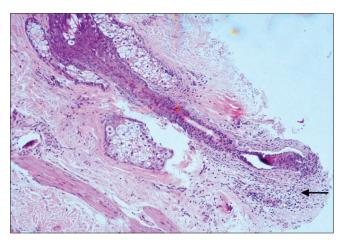


Figure 1: "Swarm of bees:" Peribulbar lymphocytic infiltrate: Anagen hair follicle [hematoxylin and eosin (×100)]

intrabulbar CD4+ T-cell infiltrate was seen in 8 (26.7%) cases, whereas a CD8+ T-cell infiltrate was seen in 21 (70%) cases [Figures 3 and 4]. From the 30 cases in our study, hair follicle dermal papilla could be identified in the section in 19 cases, out of which 7 (36.8%) cases were positive for Phosphorylated transducer and activator of transcription-1. Positivity in two other locations – terminal germinative unit and periadnexal area were seen in two additional cases. The grade of staining was faint in five cases, moderate in two cases and intense in two cases [Figure 5].

A peribulbar infiltrate was identified in 27 cases and out of these 9 (33.3%) cases were positive for Phosphorylated transducer and activator of transcription-1. None of the cases without a peribulbar infiltrate showed signal transducer and activator of transcription-1 positivity. This association was not found to be statistically significant (P value = 0.480).

Phosphorylated transducer and activator of transcription-1 positivity was seen in 4 out of 9 (44.4%) patients with a disease duration of <3 months, 2 out of 7 cases (28.5%) with a disease duration of 3–12 months and 2 out of 4 cases (50%) with a disease duration of 12–24 months, while it was seen in 1 out of 4 cases (25%) with a disease duration between 2 and 5 years, and none out of the six cases with a disease duration of >5 years. However, the association between phospho-signal transducer and activation of transcription-1 positivity with the disease duration was not found to be statistically significant (P value = 0.352) Phosphorylated transducer and activator of transcription-1 was positive in 8 out of the 22 (36.3%) alopecia areata (patchy) cases and 1 out of 6 (16.6%) alopecia universalis cases, and none of the alopecia totalis cases.

Phosphorylated transducer and activator of transcription-1 positivity was seen in 7 of 17 cases that had <25% hair loss, 1 of 4 cases with 25–50% hair loss and 1 of 8 cases with 100% hair loss. This association was not found to be statistically significant (P value = 0.534).

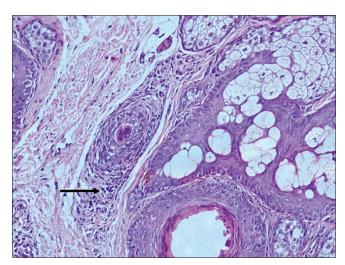


Figure 2: Peribulbar lymphocytic infiltrate around a miniaturized hair follicle (H and E, $\times 100$)

These findings have been summarized in Tables 2–4.

Discussion

Alopecia areata is a non-cicatricial alopecia caused by a T-cell-mediated autoimmune attack as a result of the immune privilege collapse of the anagen hair follicle.³ Male preponderance was observed in our study with 19 (63%) male and 11 (27%) female patients with a male to female ratio of 2.6:1, which is in accordance with the findings noted in other studies from India.^{1,11} In our study, 16 (53%) patients presented within

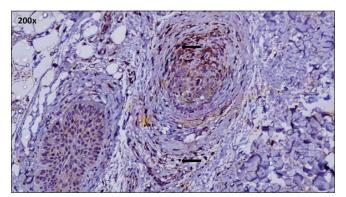


Figure 3: CD4+ T cell infiltrate: Peribulbar and intrabulbar (IHC, ×200)

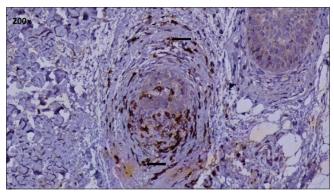


Figure 4: CD8+ T cell infiltrate: Peribulbar and intrabulbar(IHC, ×200)

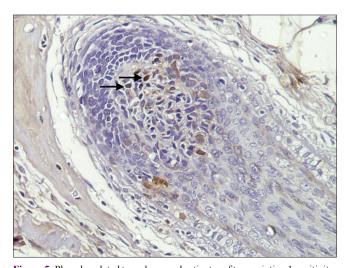


Figure 5: Phosphorylated transducer and activator of transcription 1 positivity: Hair follicle dermal papilla (IHC, ×400)

1 year of disease onset. In a study by Jain *et al.*, 78% patients presented within 6 months of disease onset. The majority of our patients (73.3%) presented with patchy disease pattern, similar to various studies that report patchy alopecia areata as the most common (75%) pattern in the world, ⁴ as well as in India. ^{11,12}

The mean follicular count was found to be 11.92 in our study, similar to findings that have been reported in various studies where the follicular count in alopecia areata has been found to be reduced by 25% from the normal count of 30–40. 13,14

The anagen hair were found to be decreased, with a mean number of 1.63. There was an increase in the nonanagen hair – catagen and telogen hair, with the anagen: telogen ratio being 1:4.1.

Vellus hair were found to be increased and terminal hair decreased, with a terminal to vellus ratio of 3.8:1. Other studies report similar findings with a decrease in the anagen

Table 2: Summary of important findings							
Parameter	Mean						
Age	24.37±14.048 years						
Follicular count	11.93±7.235						
Anagen hair	1.63±1.586						
Telogen hair	7.84±5.471						
Terminal hair	7.83±5.471						
Vellus hair	2.47±2.417						
Gender distribution	19 males and 11 females; 2.6:1						
Disease duration	30% presented within first 3 months						
Pattern	22 patients: Patchy3: Ophiasis1: Reticular						
Extent	22: AA 6: AU 2: AT						
T:V ratio	3.8:1						
A:T ratio	1:5						
Peribulbar CD4+ T-cell infiltrate	21 patients						
Peribulbar CD8+ T-cell infiltrate	25 patients						
Intrabulbar CD4+ T-cell infiltrate	8 patients						
Intrabulbar CD4+ T-cell infiltrate	21 patients						
Phosphorylated transducer and activator of transcription-1 positivity	9 patients						

Parameters	P	Degree of freedom
Phosphorylated transducer and activator of transcription-1 positivity and peribulbar infiltrate	0.480	9
Phosphorylated transducer and activator of transcription-1 positivity and disease duration	0.352	4
Phosphorylated transducer and activator of transcription-1 positivity and SALT score	0.534	3

AA: Alopecia areata, AU: Alopecia universalis, O: Onychorrhexis, T: Trachyonychia

Table 3: Demography and clinical profile of patients with phosphorylated transducer and activator of transcription-1 positivity

Age	Sex	Duration	Pattern	Extent	Nail change	Trigger	Personal history	Family history	Severity (SALT) (%)	Thyroid disease	
18	Female	2 weeks	Patchy	AA	Nil	Nil	Absent	Diabetes	<25	Absent	
27	Male	9 months	Patchy	AA	T,O	Nil	Hypothyroidism	Absent	<25	Absent	
32	Female	2 years	Patchy	AA	Nil	Nil	Absent	Absent	<25	Present	
35	Male	2 months	Patchy	AA	Nil	Nil	Absent	Absent	<25	Absent	
28	Male	3 years	Nil	AU	Nil	Nil	Absent	Absent	100	Absent	
36	Male	6 months	Patchy	AA	Nil	Nil	Absent	Absent	<25	Absent	
4	Female	1 year	Patchy	AA	T	Nil	Absent	Absent	<25	Absent	
25	Male	2 months	Patchy	AA	Nil	Nil	Absent	Atopy	26-50	Absent	
17	Male	3 weeks	Patchy	AA	Nil	Infection	Absent	Absent	<25	Absent	

AA: Alopecia areata, AU: Alopecia universalis, O: Onychorrhexis, T: Trachyonychia

Table 4: Key findings related to pSTAT-1 positivity

(a) Histopathological and immunological profile of patients with phosphorylated transducer and activator of transcription-1 positivity

Follicular count number	A (n)	A (n)	C (n)	T (n)	Terminal hair (n)	Vellus hair (<i>n</i>)	TGU (n)	Peri-vascular lymph. infiltrate	lym	ribulba phocy filtrate	tic	lympl	bulbar nocytic Itrate	STAT-1 positivity	Location
								Grade	CD4	CD8	CD4	CD8			
8	1	0	7	8	0	0	Present	2	1	2	0	2	1	1	
6	0	1	3	4	0	0	Present	1	0	1	0	1	1	1	
3	0	0	2	2	1	0	Present	2	1	2	0	1	3	1	
9	1	0	5	6	2	0	Present	1	1	2	0	1	2	1	
17	2	0	10	12	5	1	Present	1	0	2	0	1	1	1	
32	2	12	13	27	5	3	Present	2	2	2	1	2	1	1	
7	1	1	5	7	0	0	Present	1	1	1	0	1	2	2	
8	2	0	5	7	1	0	Present	2	2	2	2	2	1	1	
10	1	0	6	7	3	0	Present	3	3	3	0	3	3	3	
							Present								

A: Anagen hair, C: Catagen hair, T: Telogen hair, TGU: Terminal germinative unit, STAT-1 positivity: 1 – Faint nuclear staining, 2 – Moderate nuclear staining, 3 – Intense nuclear staining, STAT-1 location: 1 – Dermal papilla, 2 – Peri-appendegeal, 3 – Terminal germinative unit

(b) Summary of the key findings

Phosphorylated transducer and activator of transcription-1 positivity was seen in 36.8% of the patients

Phosphorylated transducer and activator of transcription-1 positivity in two novel locations - periadnexal and terminal germinative unit

All phosphorylated transducer and activator of transcription-1 + cases had a peribulbar infiltrate

Majority (44%) of phosphorylated transducer and activator of transcription-1 + cases had a disease duration of <3 months

88% of the cases with phosphorylated transducer and activator of transcription-1 positivity had patchy alopecia areata

Majority (77.7%) of the cases showing phosphorylated transducer and activator of transcription-1 positivity had <25% hair loss

count, increase in the nonanagen hair, increase in the vellus hair and a decrease or reversal of the A:T and T:V ratio. Marwah *et al.*¹⁵ report a T:V ratio of 1:3.6; Chaitra *et al.*¹⁶ report an A: T ratio of 1:1.62.

With the collapse of the immune privilege in alopecia areata, CD8+ cells enter the hair follicle, whereas CD4+ T-cells constitute the majority of lymphocytes in the perifollicular area.^{5,6}

A peribulbar lymphocytic infiltrate was identified in 27 (90%) of our cases, whereas Marwah *et al.* reported it in 76% of their cases.¹⁵

A peribulbar CD4+ T-cell infiltrate was seen in 21 (70%) cases, whereas a CD8+ T-cell infiltrate was seen in 25 (83.3%) cases. Most of the studies report that the peribulbar infiltrate is predominantly CD4+ T-cell in nature with CD4:CD8 ratios varying from 1.1:1 to 2.3:1.^{17,18} CD4:CD8 ratio has been shown to vary based on the disease activity,¹⁹ severity²⁰ and pattern,¹⁷ and the findings in our study could be because of an over-representation of severe disease in the sample because CD4:CD8 ratio decreases with increasing disease severity.²⁰ However, Xing *et al.*⁷ have reported CD8+ T-cells as the predominant cell type and found CD4 T-cells to be infrequent in alopecia areata lesional skin.

In our study, the intrabulbar infiltrate was composed of CD8+ T-cells among 21 (70%) of the cases, similar to the findings of Todes-Taylor *et al.*¹⁹ and Ranki *et al.*¹⁸

Recently, studies with mouse and human disease models have brought into picture the critical role of the CD8+ NKG2D T-cells and the interferon-y/interleukin-15 feedback loop in the pathogenesis of alopecia areata. It has been demonstrated that CD8+ NKG2D T-cells enter the dermis, surround the hair follicle bulb, binding to the major histocompatibility complex-I peptide complex and NKG2D ligand present on the hair follicle epithelial cells. These activated T-cells now produce interferon-y which binds to its receptor on the follicular epithelial cells, activates Janus kinase-1/Janus kinase-2 which causes signal transducer and activator of transcription-1 which is translocated to the nucleus leading to interleukin-15 production. Interleukin-15 now binds to its receptor complex on the CD8+ T cells, and causes Janus kinase-1/Janus kinase-3 as well as signal transducer and activation of transcription-5. enhancing the production of interferon-y, thus perpetuating this critical feedback loop [Figure 6].7 Recently, this interferony signaling through Janus kinase-1/2 has been demonstrated by the presence of phosphorylated signal transducer and activator of transcription proteins (phosphorylated transducer and activator of transcription-1, phosphorylated transducer and activator of transcription-3) in human and mouse alopecic hair follicles, but not in normal hair follicles.⁷

In our study, using immunohistochemistry we found phosphorylated transducer and activator of transcription-1 positivity in nine of our cases.

Out of the 30 vertical sections assessed, hair follicle dermal papillae were identified in 19 cases. Phosphorylated transducer and activator of transcription-1 positivity was found in the hair follicle dermal papilla cells in 7 (36.8%) cases, whereas positivity in two other novel locations – terminal germinative unit and periadnexal area was seen in one case each. The reduced hair follicle dermal papilla count in our study may be because of a 25% reduction in follicle numbers that has been reported in alopecia areata, from the normal follicular count of 30-40 hair per 4 mm biopsy. 13,14 However, these values have been reported from studies done in a Western population, and a lesser hair density has been shown among Asians.21 Furthermore, with vertical sectioning, as was done in our study, only 10-15% of the sample follicles are visualized; therefore several serial sections²² and multiple biopsies may be necessary²³ to maximize the diagnostic yield, which was not feasible in our set up owing to a limitation of resources and ethical concerns.

In a study done by Xing *et al.*, patients fulfilling the following criteria were selected: 30–95% hair loss owing to alopecia areata, hair loss duration of at least 3 months; subject age 18–75 years.⁷ Phosphorylated transducer and activator of transcription-1 positivity was seen in five patients using Western-blot analysis and percentage positivity was not elaborated. However, in our

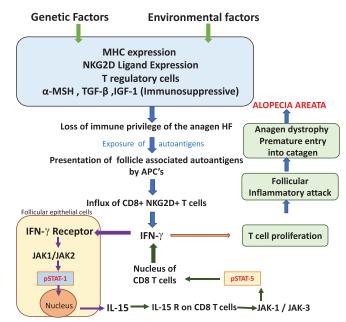


Figure 6: Critical role of the CD8+ NKG2D T cells and the interferon-γ/interleukin-15 feedback loop in the pathogenesis of alopecia areata. CD8+NKG2D T cells enter the dermis, surround the hair follicle bulb, bind to the major histocompatibility complex-I peptide complex and NKG2D ligand present on the hair follicle epithelial cells. These activated T cells now produce interferon-γ which binds to its receptor on the follicular epithelial cells, activates Janus kinase-1/Janus kinase-2 which further cause signal transducer and activator of transcription-1 activation, which is translocated to the nucleus leading to interleukin-15 production. Interleukin-15 now binds to its receptor complex on the CD8+ T-cells and causes Janus kinase-1/Janus kinase-3 as well as signal transducer and activator of transcription-5 activation, enhancing the production of interferon-γ, thus perpetuating this critical feedback loop

study in all patients of alopecia areata irrespective of age, disease duration and percentage of hair loss, an immunohistochemical analysis was done. It remains to be elucidated on the extent to which these factors might have further affected the phosphorylated transducer and activator of transcription-1 positivity. We also made an attempt to correlate the presence of phosphorylated transducer and activation of transcription-1 with the other available parameters. In our study none of the cases without a peribulbar infiltrate showed signal transducer and activator of transcription-1 positivity, but this association was not found to be statistically significant (*P* value = 0.480).

A large number of cases, i.e. 4 out of 9 cases (44%), demonstrating phosphorylated transducer and activator of transcription-1 positivity presented with an acute onset of disease (<3 months duration). In our study, 8 out of 9 (88%) cases with phosphorylated transducer and activation of transcription-1 positivity had localized disease (alopecia areata), while only one patient with alopecia universalis demonstrated phosphorylated transducer and activation of transcription-1 positivity. These findings were similar to those of Xing *et al.* where all the cases with phosphorylated transducer and activator of transcription-1 positivity were alopecia areata and no alopecia totalis or alopecia universalis case was included in the study.⁷

A majority of our patients, i.e. 7 out of 9 (77.7%), with phosphorylated transducer and activator of transcription-1 positivity had <25% hair loss, whereas only one case each belonged to the 25–50% and 100% hair loss categories. Xing *et al.* included only patients with 30–95% hair loss in their study and found no association with disease extent or severity. Thus, further research is required to study if this positivity has any association with extent of disease.⁷

Limitations

The limitations of our study, as mentioned earlier, included a small sample size, performing a single biopsy and the use of only vertical sectioning which may have adversely affected the detection of phosphorylated transducer and activator of transcription-1 positivity.

Conclusion

The present study assessed the clinical features, histopathology and immunology of alopecia areata. A peribulbar lymphocytic infiltrate was identified in majority of our cases which was composed of both CD4+ and CD8+ T-cells while the intrabulbar infiltrate was composed predominantly of CD8+ T-cells.

Phosphorylated transducer and activator of transcription-1 positivity as an indicator of Janus kinase-1/2 signaling pathway was seen in 7 (36.8%) of our cases, demonstrating the critical role the role of this pathway in the pathogenesis of alopecia areata.

Furthermore, we propose that phosphorylated transducer and activator of transcription-1 positivity may be an indicator of acute, progressive disease with an active CD8 T-cell predominant peribulbar inflammatory infiltrate indicative of a possible good treatment response.

With many recent studies suggesting that pharmacological inhibition of this pathway via Janus kinase-signal transducer and activator of transcription inhibitors such as ruxolitinib and tofacitinib leads to restoration of hair growth in alopecia areata, the understanding of this critical signaling pathway has become even more relevant today.²⁴

However, further studies with a larger sample size are needed to clearly establish the role of the Janus kinase–signal transducer and activator of transcription pathway in the pathogenesis of alopecia areata.

Declaration of patient consent

The authors certify that they have obtained all appropriate patient consent forms. In the form, the patients have given their consent for their images and other clinical information to be reported in the journal. The patients understand that name and initials will not be published and due efforts will be made to conceal identity, but anonymity cannot be guaranteed.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

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